



Determination of Polycyclic Aromatic Hydrocarbons (PAHs) in Seafood Using GC/MS

UCT Part Numbers:

ECQUUS2-MP (4 g of muffled anh. MgSO₄ and 2 g of NaCl)

ECPAHFR50CT (50 mL centrifuge tubes, PAHs removed)

EUSILMSSM26 (6 mL, 1g silica gel cartridge with 200 mg of muffled anhydrous sodium sulfate on top)

Background

This method is used for the determination of polycyclic aromatic hydrocarbons (PAHs), polychlorinated biphenyls (PCBs) and polybrominated diphenyl ethers (PBDEs) in fish and seafood--oyster, shrimp, and mussel. Benzo[a]pyrene is the main analyte of interest. GC/MS instrumentation is used for analysis.

PAH Analytes Covered in this Method

| PAH | Abbreviation | PAH | Abbreviation |
|------------------------------|--------------|--------------------------------------|--------------|
| Anthracene | Ant | Indeno[1,2,3-<i>cd</i>]pyrene | IcdP |
| Benz[a]anthracene | BaA | Naphthalene | Naph |
| Benzo[a] pyrene | BaP | Phenanthrene | Phe |
| Benzo[b]fluoranthene | BpF | Pyrene | Pyr |
| Benzo[k]fluoranthene | BkF | 3-Methylchrysene | 3-MC |
| Benzo[g,h,i]perylene | BghiP | 1-Methylnaphthalene | 1-MN |
| Chrysene | Chr | 1-Methylphenanthrene | 1-MP |
| Dibenz[a,h]anthracene | DBahA | 2,6 Dimethylnaphthalene | 2,6-DMN |
| Fluoranthene | Flt | 1,7-Dimethylphenanthrene | 1,7-DMP |
| Fluorene | Fln | | |

Procedure

1) Extraction

- a) To the 50-mL polypropylene centrifuge tube add 10 ± 0.1 g of homogenized seafood sample
- b) Add 50 μ L of 1 μ g/mL ^{13}C -PAHs solution
- c) Vortex sample for 15 sec and then equilibrate for 15 min
- d) Add 5 mL of reagent water and 10 mL of ethyl acetate (EtOAC)
- e) Shake for 1 min

2) Partition

- a) Add the contents of pouch **ECQUUS2-MP**. Tightly seal the tube to ensure that salts do not get into the screw threads
- b) Shake for 1 min
- c) Centrifuge at $> 1,500$ rcf for 10 min
- d) Remove 5-mL aliquot of the upper ethyl acetate layer, add 50 μ L of isooctane as a keeper
- e) Evaporate all ethyl acetate until only isooctane and co-extracted sample fat remain
- f) Reconstitute with 1 mL of hexane

3) Clean-Up

- a. Condition a silica SPE column **EUSILMSSM26 (Note 1)** (1 g of silica gel with approx. 0.2 g of muffled anh. sodium sulfate on the top) with 6 mL of hexane:dichloromethane (3:1 v/v) and 4 mL of hexane
- b) Apply the reconstituted extract to the silica SPE cartridge (**Note 2**)
- c) Elute with 10 mL of hexane:dichloromethane (3:1 v/v) and collect the eluent
- d) Add 0.5 mL isooctane to the eluent as a keeper and evaporate to 0.5 mL to remove hexane and dichloromethane from the final extract
- e) Transfer the final extract into an autosampler vial for GC/MS analysis

Notes:

1. The fat capacity of the 1-g silica gel SPE column is approx. 0.1 g. If the ethyl acetate extract aliquot contains more than 0.1 g of fat, use a smaller aliquot to avoid sample breakthrough
2. Ethyl acetate should not be present in the extract applied to the silica cartridge as it affects extract polarity and potential retention of fat and analytes on the silica gel.

GC Conditions for the Analysis of PAHs

| | |
|----------------------------------|---|
| Column | BPX-50 (30 m x 0.25 mm i.d. x 0.25 µm film thickness) |
| Oven Temperature Program | 80°C (hold for 4.3 min), 30°C/min to 220°C, 2°C/min to 240°C, and 10°C/min to 360°C (hold for 17 min) |
| He Flow Rate | 1.3 mL/min (hold for 19 min), then 50 mL/min to 2 mL/min (hold for 16 min) |
| Injection Technique | PTV solvent vent |
| Injection Volume | 1 x 8 µL |
| Vent Time | 2.3 min |
| Vent Flow | 50 mL/min |
| Vent Pressure | 50 psi |
| Inlet Temperature Program | 50°C (hold for 2.3 min), then 400°C/min. to 300°C |

MS Conditions

Any GC-MS instrument (single quadrupole, triple quadrupole, time-of-flight or ion trap) with electron ionization (EI) may be used

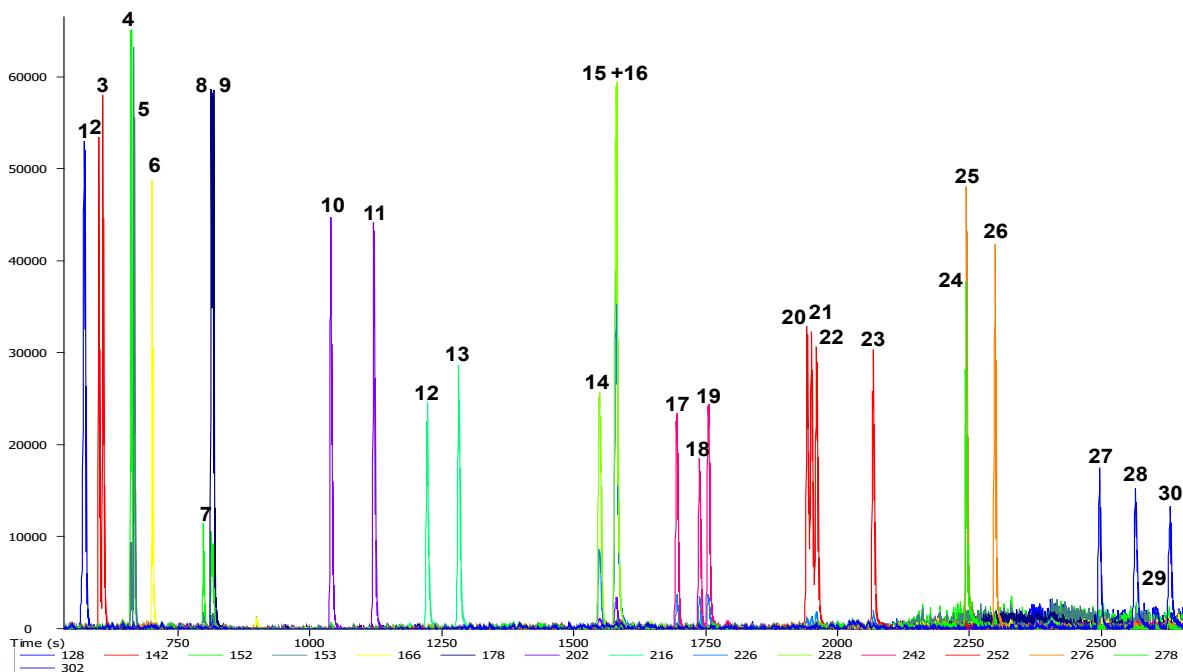
**MS Ions (*m/z*) for Quantification and Identification of Target PAHs
for Single-stage MS Instruments**

| Analyte PAH's | Abbreviation | Confirmation Ions (<i>m/z</i>) | Quantification Ions (<i>m/z</i>) |
|---------------------------------|--------------|-------------------------------------|---------------------------------------|
| Anthracene | Ant | 177 | 178 |
| Benz[a]anthracene | BaA | 226 | 228 |
| Benzo[a] pyrene | BaP | 253 | 252 |
| Benzo[b]fluoranthene | BpF | 253 | 252 |
| Benzo[k]fluoranthene | BkF | 253 | 252 |
| Benzo[g,h,i]perylene | BghiP | 277 | 276 |
| Chrysene | Chr | 226 | 228 |
| Dibenz[a,h]anthracene | DBahA | 276 | 278 |
| Fluoranthene | Flt | 200 | 202 |
| Fluorene | Fln | 165 | 166 |
| Indeno[1,2,3-cd]pyrene | IcdP | 277 | 276 |
| Naphthalene | Naph | 127 | 128 |
| Phenanthrene | Phe | 177 | 178 |
| Pyrene | Pyr | 200 | 202 |
| 3-Methylchrysene | 3-MC | 241 | 242 |
| 1-Methylnaphthalene | 1-MN | 115 | 142 |
| 1-Methylphenanthrene | 1-MP | 189 | 192 |
| 2,6 Dimethylnaphthalene | 2,6-DMN | 141 | 192156 |
| 1,7-Dimethylphenanthrene | 1,7-DMP | 191 | 156206 |

**MS Ions (*m/z*) for Quantification and Identification of Target ^{13}C -PAHs
for Single-stage MS Instruments**

| Analyte PAH's | Abbreviation | Confirmation Ions (<i>m/z</i>) | Quantification Ions (<i>m/z</i>) |
|---|-----------------------------|-------------------------------------|---------------------------------------|
| Naphthalene ($^{13}\text{C}_6$) | Naph- $^{13}\text{C}_6$ | 133 | 134 |
| Fluorene ($^{13}\text{C}_6$) | Fln- $^{13}\text{C}_6$ | 171 | 172 |
| Phenanthrene ($^{13}\text{C}_6$) | Phe- $^{13}\text{C}_6$ | 183 | 184 |
| Anthracene ($^{13}\text{C}_6$) | Ant- $^{13}\text{C}_6$ | 183 | 184 |
| Fluoranthene ($^{13}\text{C}_6$) | Flt- $^{13}\text{C}_6$ | 205 | 208 |
| Pyrene ($^{13}\text{C}_6$) | Pyr- $^{13}\text{C}_6$ | 208 | 205 |
| Benz[a]anthracene ($^{13}\text{C}_6$) | BaA- $^{13}\text{C}_6$ | 232 | 234 |
| Chrysene ($^{13}\text{C}_6$) | Chr- $^{13}\text{C}_6$ | 232 | 234 |
| Benzo[b]fluoranthene ($^{13}\text{C}_6$) | BbF- $^{13}\text{C}_6$ | 259 | 258 |
| Benzo[k]fluoranthene ($^{13}\text{C}_6$) | BkF- $^{13}\text{C}_6$ | 259 | 258 |
| Benzo[a]pyrene ($^{13}\text{C}_4$) | BaP- $^{13}\text{C}_4$ | 257 | 256 |
| Indeno[1,2,3-<i>cd</i>]pyrene ($^{13}\text{C}_6$) | IcdP- $^{13}\text{C}_6$ | 283 | 282 |
| Dibenz[a,h]anthracene ($^{13}\text{C}_6$) | DBahA- $^{13}\text{C}_6$ | 282 | 284 |
| Benzo[g,h,i]perylene ($^{13}\text{C}_{12}$) | BghiP- $^{13}\text{C}_{12}$ | 289 | 288 |

An example chromatogram of a GC separation of PAHs and their alkyl homologues in a standard solution mixture at 25 ng/mL in isoctane



1 – naphthalene, 2 – 2-methylnaphthalene, 3 – 1-methylnaphthalene, 4 – acenaphthylene,
5 – acenaphthene, 6 – fluorene, 7 – dibenzothiophene, 8 – phenanthrene, 9 – anthracene,
10 – fluoranthene, 11 – pyrene, 12 – 1-methylpyrene, 13 – benzo[c]fluorene,
14 – benz[a]anthracene, 15 – cyclopenta[c,d]pyrene, 16 – chrysene, 17 – 1-methylchrysene,
18 – 5-methylchrysene, 19 – 3-methylchrysene, 20 – benzo[b]fluoranthene,
21 – benzo[k]fluoranthene, 22 – benzo[j]fluoranthene, 23 – benzo[a]pyrene,
24 – dibenz[a,h]anthracene, 25 – indeno[1,2,3-cd]pyrene, 26 – benzo[g,h,i]perylene,
27 – dibenzo[a,l]pyrene, 28 – dibenzo[a,e]pyrene, 29 – dibenzo[a,h]pyrene,
30 – dibenzo[a,l]pyrene

*The analyst should refer to Katerina Mastovska, Wendy R. Sorenson, Covance Laboratories Inc
Jana Hajslova, Institute of Chemical Technology, Prague "Determination of Polycyclic Aromatic Hydrocarbons (PAHs) in Seafood using Gas Chromatography-Mass Spectrometry: A Collaborative Study"

References

Lucie Drabova, Kamila Kalachova, Jana Pulkarbova, Tomas Cajka, Vladimir Kocourek and Jana Hajslova. "Rapid Method for Simultaneous Determination of Polycyclic Aromatic Hydrocarbons (PAHs), Polychlorinated Biphenyls (PCBs) and Polybrominated Diphenyl Ethers (PBDEs) in Fish and Sea Food Using GC-TOFMS," ICT document, Prague, Czech Republic, 2010.

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